

Table 1.

	GG	GT	TT
AA	3	4	9
AT	2	6	0
TT	6	0	0

Table 2.

Site 218	Site 243	Frequency
T	A	0.4
G	T	0.4
G	A	0.2
T	T	0

Chromosome location: The location of the canine gene is unknown; the corresponding human gene maps to 17q21 (Law *et al.* 1986), or to 17q11.1-q12 (Xu *et al.* 1988).

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References

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A polymorphism in intron D of the chinook salmon growth hormone 2 gene

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Source/description: Locus-specific primers were developed using published gene and cDNA sequences of growth hormone 1 and growth hormone 2 from *Salmo salar*, *Oncorhynchus mykiss*, and *Oncorhynchus keta* (Agellon & Chen 1988; Agellon *et al.* 1988; Johansen *et al.* 1989; Sekine *et al.* 1989).

Primer sequences:

Forward primer (GH2ex-4): 5'-CAG CCT AAT GGT CAG AAA CT-3'
 Reverse primer (GH2ex-5a): 5'-CGT AGT TCC TCC TGA CGT TG-3'
 Expected product size: approximately 1300 base pairs.

PCR/RFLP conditions: 30 µl reactions consisting of: 0.2–0.5 µg total genomic DNA; 0.1 µM each primer; 0.2 µM dNTPs; 3 mM MgCl₂; 50 mM KCl; 10 mM Tris-HCl; 0.1% Triton; 0.3 U Taq polymerase. Amplification was performed using a profile of 1 min at 94°C, 2 min at 52°C, and 1 min at 72°C for 40 cycles, with a final extension of 10 min at 72°C. Restrictions were performed according to manufacturer's instructions and the resulting fragments were separated by electrophoresis on a 1.5% agarose gel, and visualized using ethidium bromide and UV light.

Polymorphism: RFLPs were detected in DNA samples from four populations of chinook salmon in the Columbia River basin (northwestern USA) using the endonuclease *DraI*. Allele A contains two *DraI* sites, which yield fragments of approximately 550, 520, and 230 bp. Allele B contains only a single *DraI* site and yields two fragments of approximately 1070 and 230 bp. Restriction with the

enzymes *BamHI*, *EcoRI*, *PstI*, and *HaeIII* revealed no polymorphisms. Restriction with the enzyme *BsrGI* revealed a polymorphism linked to the *DraI* polymorphism and, therefore, yielded no additional information.

Mendelian inheritance: Segregation of the alleles was examined in two heterozygote–homozygote crosses. A sample of 15 offspring of undetermined sex from each of the two crosses (see Fig. 1 and Table 1) showed codominant segregation of the two alleles. No significant departures from expected values were observed ($\chi^2 = 0.6$ for each cross), although the power of the test was limited by the small sample size.

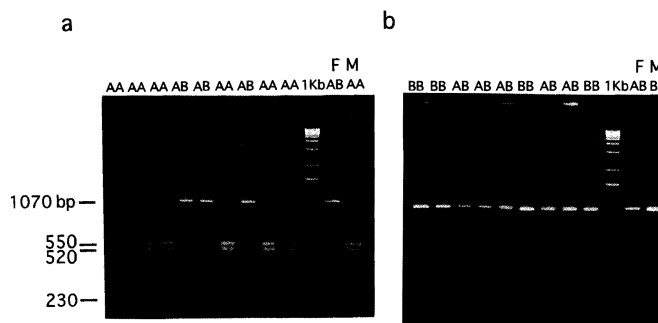


Fig. 1. Growth hormone 2 intron D cut with *DraI*; nine chinook salmon progeny and parents (parents are last two lanes on gel). (a) AB female × AA male; (b) AB female × BB male.

Table 1. Genotype frequencies for the growth hormone 2, intron D polymorphism in offspring of two separate crosses of chinook salmon from Abernathy Hatchery, WA, USA

Female × Male	Offspring			Total
	AA	AB	BB	
AB × AA	6	9	0	15
AB × BB	0	6	9	15

Allele frequency: The frequencies of RFLPs were determined in a total of 68 individuals from four populations in the Columbia River drainage (Table 2).

Table 2. Frequency of growth hormone 2, intron D *DraI* polymorphism in four populations of chinook salmon from the Columbia River drainage (northwestern USA)

Location	Genotype frequency				Allele frequency	
	AA	AB	BB	Total	A	B
Lostine River, OR	15	5	0	20	0.88	0.12
Minam River, OR	17	3	0	20	0.93	0.07
Carson Hatchery, WA	10	4	0	14	0.86	0.14
Abernathy Hatchery, WA	8	5	1	14	0.75	0.25

Chromosomal location: Unknown, autosomal.

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